

Illumina sequencing technology application to Infectious Disease Surveillance

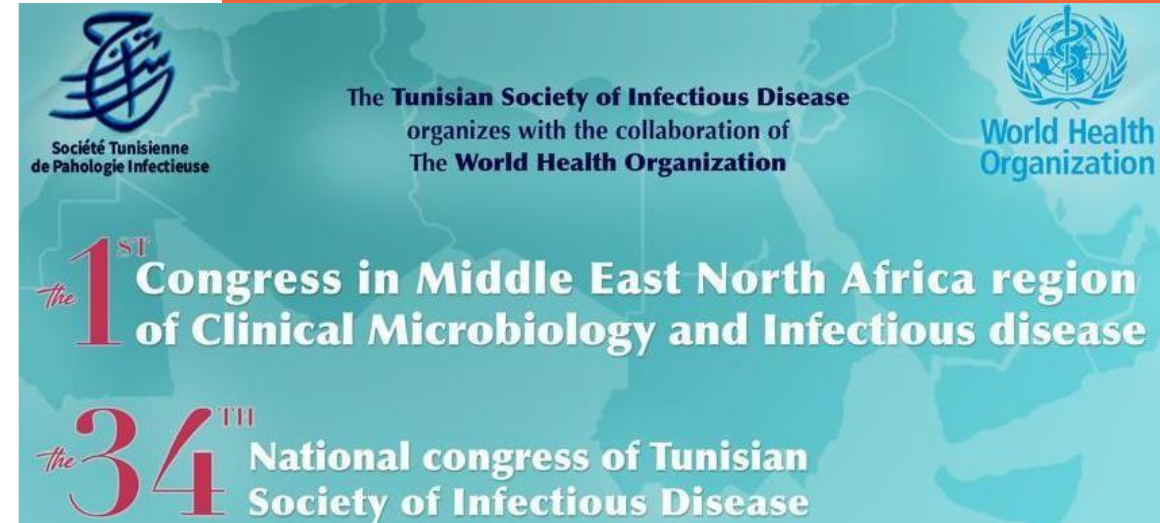
Lydia Hadjeras, PhD

Genomic specialist at Illumina

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Agenda

1 **Genomic utility in enhanced infectious diseases surveillance & AMR profiling**

2 **Illumina NGS solutions for infectious diseases surveillance & Microbiology**

Genomic utility in enhanced infectious disease surveillance & AMR profiling

1

Infectious Diseases by the Numbers

| | |
|----------|---|
| 770M | Cumulative cases of COVID-19 ³ |
| 400K | Respiratory deaths each year caused by influenza virus ⁴ |
| 10.6M | People that fell ill from tuberculosis in 2021 ⁵ |
| 39M | People living with HIV in 2022 ⁶ |
| 3.8% | Global GDP lost due to antimicrobial resistance by 2050 ⁷ |
| 247M | Malaria cases in 84 malaria-endemic countries in 2021 ⁵ |
| 1.5M | New hepatitis B infections globally in 2019 ⁵ |
| \$28-45B | Direct annual cost of treating healthcare associated infections in the US ⁸ |

3. <https://covid19.who.int/> (Accessed Sept. 7, 2023); 4. <https://ourworldindata.org/influenza>; 5. World health statistics 2023: monitoring health for the SDGs, Sustainable Development Goals. Geneva: World Health Organization; 2023.; 6. <https://www.unaids.org/en/resources/fact-sheet>; 7. Global antimicrobial resistance and use surveillance system (GLASS) report 2022. Geneva: World Health Organization; 2022.; 8. Gidey K, et al. Clinical and economic burden of healthcare-associated infections: A prospective cohort study. PLoS One. 2023 Feb 23;18(2):e0282141.



Infectious
diseases impact
human health
and economics

A man wearing a black cap, sunglasses, and a grey long-sleeved shirt is feeding a large number of brown chickens in a multi-tiered industrial farm cage. He is holding a red bowl and pouring feed into a trough. The background shows the structure of the farm with wooden slats and metal bars.

The increasing threat of infectious diseases

1. **Globalization and increased international travel** facilitate the rapid spread of infectious agents across borders.
2. **Expanding urbanization and increasing population density** create ideal conditions for the transmission of infectious agents.
3. **Emergence of drug-resistant strains of bacteria and viruses** poses a significant challenge in the treatment and control of infections.
4. **Closer contact with animal reservoirs of infectious diseases** due to factors such as climate change, deforestation, and high-intensity farming, can increase the risk of animal-to-human transmission.⁹



Beyond COVID:

**Lessons learned from
a global pandemic**

The pandemic showcased effectiveness in public health surveillance:

1. Rapidly characterize a novel infectious agent
2. Monitor the spread of a global disease
3. Facilitate research that ultimately led to an effective vaccine in record time

“ Genomic surveillance is transforming public health action by providing a deeper understanding of pathogens, their evolution and circulation”

World Health Organization¹⁹

Antimicrobial Resistance (AMR)

An Important Target for Surveillance

- WHO declared AMR “One of the top 10 global public health threats facing humanity”¹
- AMR complicates the treatment of microbial infections and can lead to multi-drug resistant organisms
- Currently majority of AMR surveillance relies on commonly available microbiological methods and phenotypic drug susceptibility testing, with some molecular testing
- NGS can bring a significant improvement in AMR characterization
 - Quicker more accurate outbreak identification
 - Help guide development of new treatments
 - Identification of comprehensive AMR in multi-drug resistant organisms such as tuberculosis²

The One Health Approach

NGS-based infectious disease surveillance has already supported several public health initiatives, including **One Health** – an interconnective approach that promotes human health by ensuring the health of animals and plants within the shared environment.²¹



NGS allows to monitor threats to both humans and animals, including:

- Mobile genetic elements encoding antimicrobial resistance (AMR).
- Genetic mutations associated with emerging strains of infectious agents.
- Genomic surveillance of environmental samples, such as wastewater.

Illumina NGS solutions for infectious disease surveillance & Microbiology

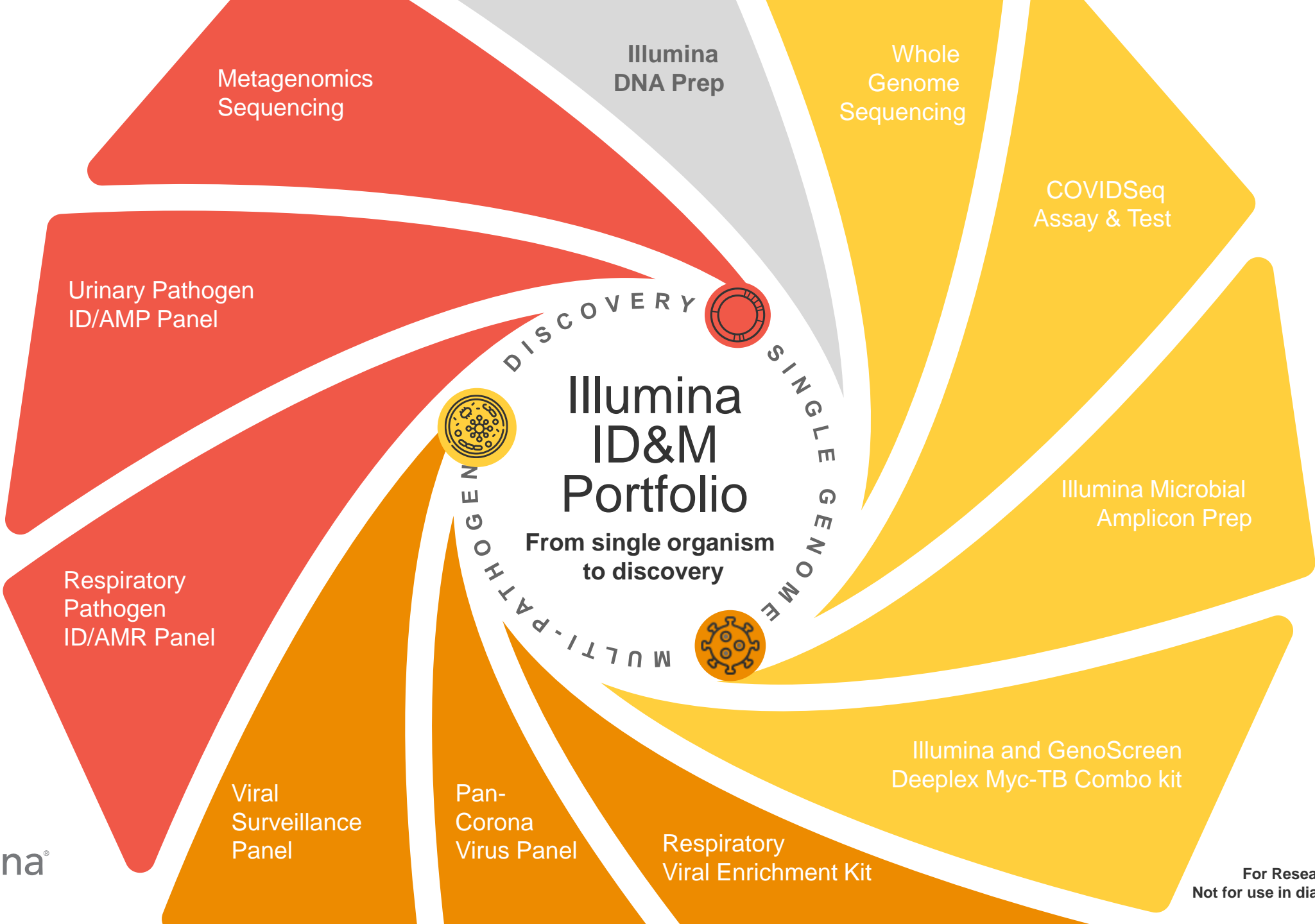
2

Illumina technologies – a unique advantage against infectious diseases



Flexible and scalable solutions to enable accessibility worldwide

- ✓ Novel pathogen ID and drug resistance characterization
- ✓ Insights for clinical assay development
- ✓ Pathogen monitoring and genomic surveillance
- ✓ Host genetics and immune response
- ✓ Drug and vaccine development



NGS Methods Enable Genomic Investigation



Whole-Genome Sequencing of Isolates

Metagenomic or Metatranscriptomic

Hybrid Capture Enrichment

Amplicon

- Analyze full microbial and plasmid genomes from cultured isolate
- **Discover new variants**
- Study anti-microbial resistance, virulence, or other **functional gene analysis**

- **Culture independent method**
- Analyze all microbes, genes, or gene expression present in complex samples
- **Analyze gene expression data**
- Can require high read-depth

- **Culture independent method**
- Can provide whole genome results
- Enables enrichment and **analysis of large numbers of organisms** with greater sensitivity than shotgun metagenomics

- **Culture-independent method**
- Can provide whole genome results
- Focused streamlined analysis
- **Fast and inexpensive**
- Risk of mutations affecting primer binding



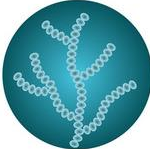

Illumina Viral Surveillance Panel (VSP)

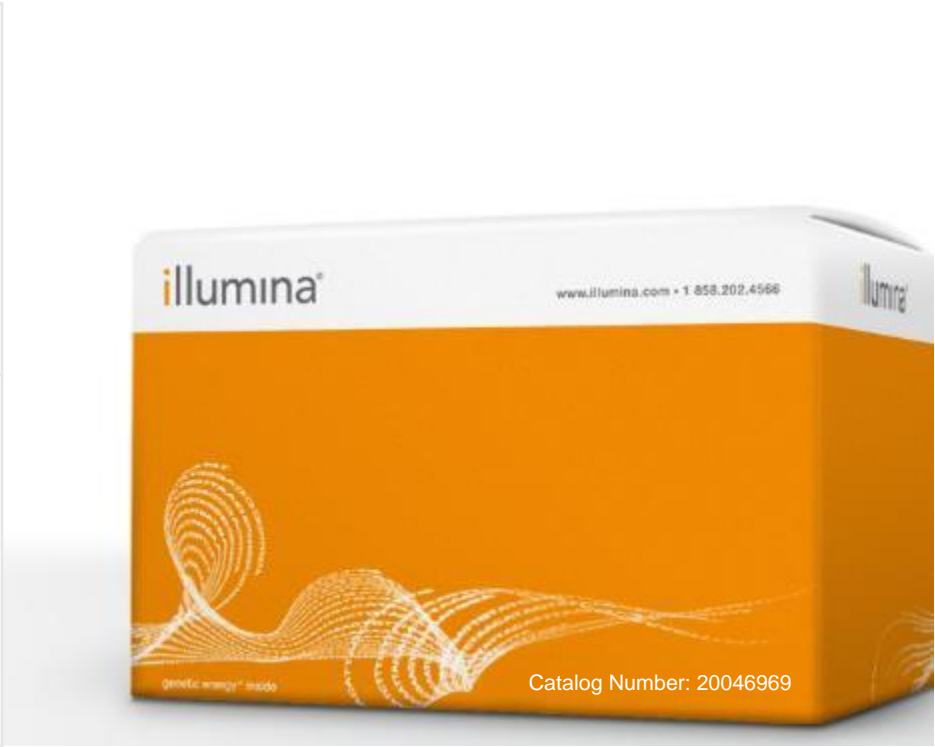
Enables Whole genome sequencing (WGS) with targeted enrichment of 200+ viruses of high risk to public health¹



| Arthropod transmitted/ febrile tropical | Bloodborne | Encephalitis | Respiratory/ Cardiopulmonary | Hemorrhagic fever | Enteric | Oncolytic | Rash/Lesion |
|--|--------------------|---------------------------|---------------------------------|------------------------|-------------------|-------------------|--------------------|
| Bourbon | Adeno-associated 2 | Cache Valley | Adenovirus A -G | Aigai | Aichi | Epstein-Barr | Herpes Simplex 1-2 |
| Chikungunya | Hepatitis A | CA encephalitis | Bocavirus | Bombali | Coxsackieviruses | Cytomegalovirus | Measles |
| Colorado Tick Fever | Hepatitis B | Cedar valley | Coronavirus (4) | Bundibugyo | Echovirus | HPV (28) | Menangle |
| Dengue 1 - 4 | Hepatitis C | Eastern equine enceph | Hantaviruses (15) | Chapare | Enterovirus A – D | Polyomavirus (13) | Mpox |
| Heartland | Hepatitis D | Ghana | Influenza A (22) | CCHF | Mamastrovirus (4) | | Mumps |
| Mayaro | Hepatitis E | Hendra | Influenza B (3) | Ebola | Norovirus | | Rubella |
| Onyong-nyong | HIV 1 - 2 | Henipah | Influenza C | Guanarito | Poliovirus | | Varicella-zoster |
| Oropuche | Torque Teno | Jamestown Canyon | MERS | Hantaviruses (7) | Rotavirus (4) | | Variola |
| Punta Toro | | Japanese enceph | Metapneumovirus | Junin | Salivirus | | |
| Ross River | | La Crosse | Parainfluenza 1 – 4 | Kyasanur Forsest | Sapovirus | | |
| Sandfly fever | | Langya | Parechovirus | Lassa | | | |
| Semliki Forest | | Lyssavirus (6) | Parvovirus B19 | Lloviu | | | |
| Sindbis | | Mojiang | RSV A | Lujo | | | |
| Tacheng tick 2 | | Murray Valley enceph. | RSV B | Machupo | | | |
| Usutu | | Nipah | Rhinovirus A – C | Marburg | | | |
| West Nile | | Powassan | SARS | Omsk hemorrhagic fever | | | |
| Yellow Fever | | Rabies | SARS-CoV-2 | Ravn | | | |
| Zika | | Snowshoe hare | | Reston | | | |
| | | St. Louise enceph. | | Rift Valley | | | |
| | | Tahyna | | Sabia | | | |
| | | Tickborne enceph. | | SFTSV | | | |
| | | Toscana | | Sosuga | | | |
| | | Venezuelan equine enceph. | | Sudan | | | |
| | | Western equine enceph | | Tai Forest | | | |

Respiratory Pathogen ID/AMR (RPIP) Enrichment Kit for Comprehensive Detection of Respiratory Pathogens and Antimicrobial Resistance Genes

| | | | |
|------------------------|---|---|--|
| >280 Pathogens |  180+ Bacteria |  40+ Viruses* |  50+ Fungi |
| >16 Antibiotic Classes |  1200+ Antimicrobial resistant alleles | | |



*full genome coverage available for SARS-CoV-2 and flu viruses

Deeplex® Myc-TB

Rapid, Culture-free, Targeted Sequencing for *M. tuberculosis* Drug Resistance Profiling



ILLUMINA AND GENOSCREEN partner to help
ELIMINATE TUBERCULOSIS
WORLDWIDE BY 2035



10 MILLION

People developed TB
worldwide in 2019



500,000

People with treatment-
resistant TB



4 in 10

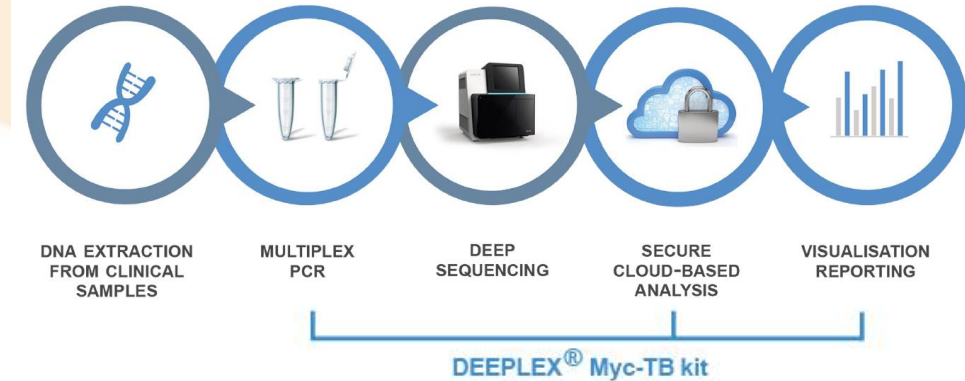
People with drug-resistant TB
diagnosed and treated

By globalizing access to the GenoScreen Deeplex®Myc-TB assay, a next-generation sequencing based assay for rapid detection of tuberculosis drug resistance, this partnership advances the World Health Organization's strategy to end the global TB epidemic by 2035.

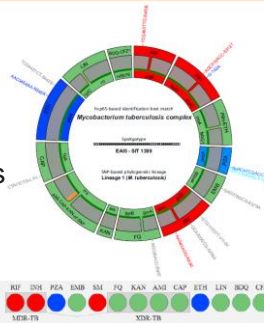
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Source: World Health Organization Global Tuberculosis Report, 2020

GenoScreen



- An all-in-one assay based on targeted sequencing
- Prediction of Resistance to 15 anti-TB drugs
- Genotyping and spoligotyping of MTBC strains
- Identification of more than 140 mycobacterial species
- Turnaround time of less than 48 hours
- Highly sensitive



WHO Rapid Communication: Use of Targeted Next Generation Sequencing (NGS) to detect DR-TB



WHO has provided its first ever guidance on the use of NGS for clinical use



Targeted next-generation sequencing (tNGS)

- *Accurate*
- *Cost-effective depending on context*
- *Acceptable and implementable under routine conditions*



World Health Organization

Use of targeted next-generation sequencing to detect drug-resistant tuberculosis

Rapid communication, July 2023

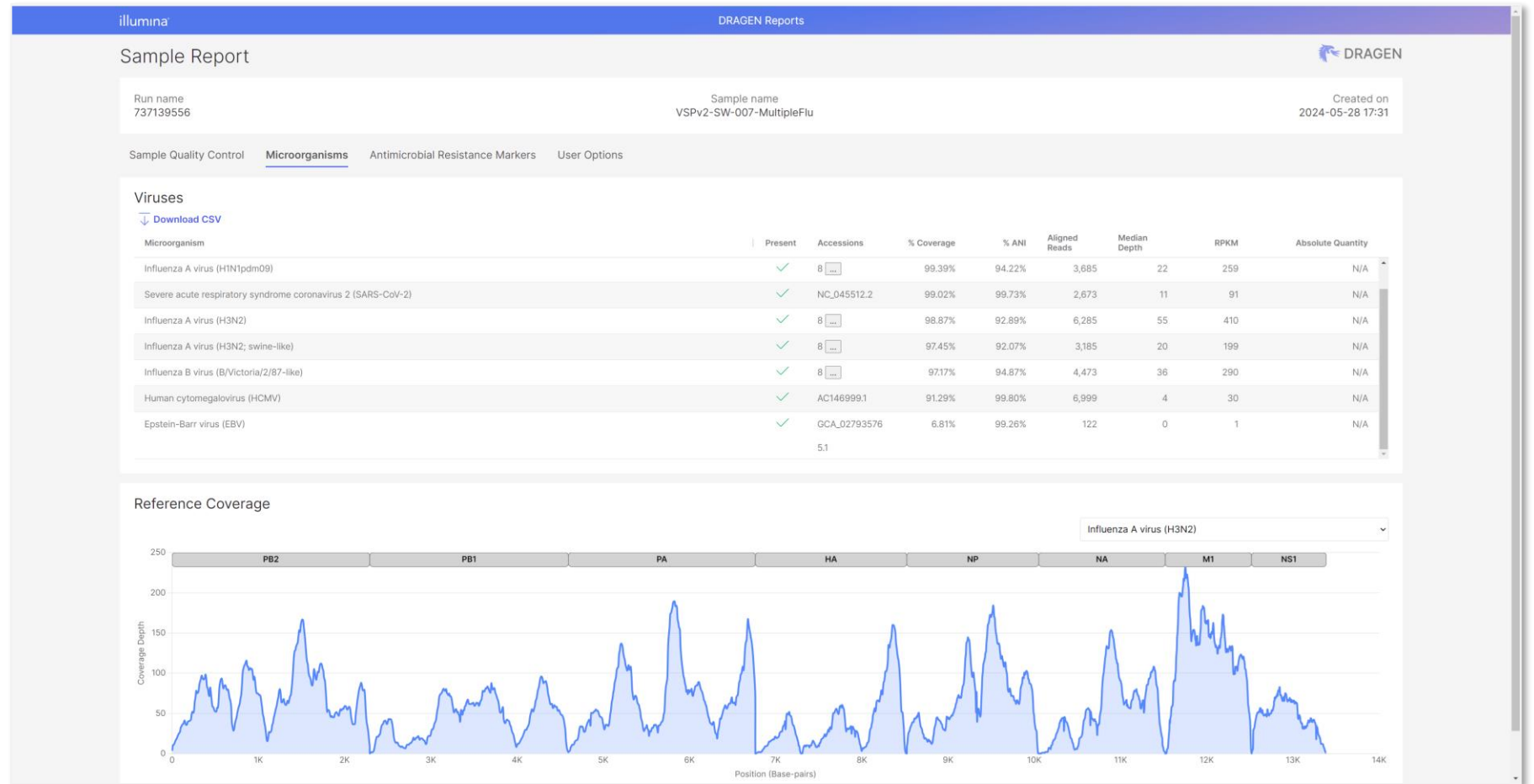
Goal: inform national TB programs and other stakeholders of key changes in the available options for detection of TB drug resistance and to allow for rapid transition and planning at the country level.

WHO Rapid Communication: Use of targeted next-generation sequencing to detect drug-resistant tuberculosis. July 2023 <https://www.who.int/publications/i/item/9789240076372>

On board DRAGEN .html report



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Example report from **DRAGEN Microbial Enrichment Plus:**

- .html dynamic report with microbial identification, coverage metrics and quantity (if using internal controls)
- Coverage plots for each genome with segments/genes (for segmented viruses)
- Each report includes QC metrics
- Delivers sufficient microbial data for quick, actionable decisions by public health labs and organizations

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Examples of how NGS methods have been applied across various infectious disease surveillance projects

Early detection

Outbreak response

Clinical research



Characterizing new and emerging pathogens

Identification of the SARS-CoV-2 virus

Conducted by:

Shanghai Public Health Clinical Center,
The Central Hospital of Wuhan

Funded by:

Center for Disease Control and Prevention (China),
National Institute for Communicable Disease Control
and Prevention (China)

NGS methodology used:

Metatranscriptomic RNA sequencing

Sequencing platform:

MiniSeq™ System

After molecular tests ruled out other etiological agents, NGS analysis of a bronchoalveolar lavage fluid sample isolated from a patient in Wuhan, China suffering from severe respiratory disease identified a novel coronavirus in December 2019.

Originally named “WH-Human 1” coronavirus after initial phylogenetic analysis revealed the virus was most closely related to a group of SARS-like coronaviruses previously identified in Chinese bats.

Rapid identification of this causative agent, later named SARS-CoV-2, marked the start of the COVID-19 pandemic.³⁰

Sequencing Enables Genomic AMR Surveillance in Wastewater and Wildlife

Sequencing provides rich information on AMR environmental transmission through wastewater AMR surveillance

- **Metagenomics** is an improved surveillance approach that could potentially limit the global burden of **AMR**
- **Metagenomic analysis of sewage** is an ethically acceptable and economically feasible approach for continuous global surveillance and prediction of AMR
- Globally applying **NGS** to human sewage will be helpful for large scale AMR surveillance
- Sequencing and bioinformatics methods are easily standardized in a central facility

Using sewage for surveillance of antimicrobial resistance
BY FRANK M. AARESTRUP, MARK E. J. WOOLHOUSE | SCIENCE 07 FEB 2020 : 630-632

POLICY FORUM | GLOBAL HEALTH

Using sewage for surveillance of antimicrobial resistance

Frank M. Aarestrup¹, Mark E. J. Woolhouse²
+ See all authors and affiliations



Metagenomics will give a complete picture of the AMR pathogen transmission in wildlife

- Successful AMR pathogens transmit between wildlife, humans, and companion animals
- The role of globally moving wildlife in the dissemination of AMR bacteria over long distances remains unclear
- **Metagenomics** can identify the abundance of AMR bacterial genes and mobile genetic elements in **the gut of wild animals**
- Continuous large-scale surveillance with metagenomics approach can provide a more complete picture of global AMR transmission in future

Wildlife Is Overlooked in the Epidemiology of Medically Important Antibiotic-Resistant Bacteria
Monika Dolejska, Ivan Literak | Antimicrobial Agents and Chemotherapy Jul 2019, 63 (8) e01167-19; DOI: 10.1128/AAC.01167-19

Wildlife Is Overlooked in the Epidemiology of Medically Important Antibiotic-Resistant Bacteria

Monika Dolejska,^{a,b} Ivan Literak^{a,b}



Antimicrobial Agents
and Chemotherapy®



Transmission Tracking

Genomic investigation of two concurrent outbreaks of *Salmonella Enteritidis* in South Africa

Conducted by:

Centre for Enteric Diseases, National Institute for Communicable Diseases (Johannesburg, South Africa)

Supported by:

Centres for Disease Control and Prevention (Africa), Centre for Enteric Diseases (South Africa), the National Institute for Communicable Diseases (South Africa)

NGS methodology used: **WGS of isolates**

Sequencing platform: **MiSeq™ System**

Outbreak A

children at a day-care center

Outbreak B

adults who all ate at the same restaurant

WGS analysis of bacterial isolates from patients affected by the outbreaks revealed that contaminated eggs from the restaurant were the likely source of the *Salmonella Enteritidis* that caused Outbreak B.

Comparative analysis of the genomes of the *Salmonella Enteritidis* strains that caused Outbreak A and Outbreak B revealed high genetic similarity, suggesting a high probability of epidemiological relatedness.

This finding indicated the **likelihood of a larger contamination problem** with the same clone of *Salmonella Enteritidis* within the wider chicken- and egg-producing industry of South Africa.³⁸

Hendra virus surveillance in Australian horses for the detection of novel variants

Genomic surveillance for the highly pathogenic zoonotic Hendra virus (HeV), which naturally infects bats, has identified several spillover events during which HeV was transmitted from bats to horses.

Although horse-to-human transmission of HeV is very rare (only seven human cases have been reported), the fatality rate in humans is extremely high at 57%.

Horse HeV vaccine developed (Equivac HeV) to prevent horse-to-human transmission² as well as monoclonal antibodies for emergency post-exposure prophylaxis

September 2020, an Australian horse exhibiting HeV symptoms was humanely euthanized and tested for HeV. However, routine quantitative RT-PCR and ELISA testing did not identify HeV genomic material or proteins, respectively.

Through **metagenomic sequencing**, researchers identified a novel variant of HeV that evaded detection by routine sentinel surveillance.

- 1- Marsh GA et al. Genome sequence conservation of Hendra virus isolates during spillover to horses, Australia. Emerging Infect Dis. 2010;16(11):1767-1769. doi:10.3201/eid1611.100501
- 2- Annand EJ et al. Novel hendra virus variant detected by sentinel surveillance of horses in australia. Emerging Infect Dis. 2022;28(3):693-704. doi:10.3201/eid2803.211245

NGS supports the need for updated routine molecular testing used in surveillance and exemplifies the power of genomic surveillance.



Merci

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